



## Production of $^{15}\text{N}$ -labeled $\alpha$ -amanitin in *Galerina marginata*



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### ARTICLE INFO

#### Article history:

Received 11 May 2015

Received in revised form

15 June 2015

Accepted 17 June 2015

Available online 19 June 2015

#### Keywords:

Amanita

Galerina

$\alpha$ -Amanitin

Isotope labeling

LC-MS

NMR

### ABSTRACT

$\alpha$ -Amanitin is the major causal constituent of deadly *Amanita* mushrooms that account for the majority of fatal mushroom poisonings worldwide. It is also an important biochemical tool for the study of its target, RNA polymerase II. The commercial supply of this bicyclic peptide comes from *Amanita phalloides*, the death cap mushroom, which is collected from the wild. Isotopically labeled amanitin could be useful for clinical and forensic applications, but  $\alpha$ -amanitin has not been chemically synthesized and *A. phalloides* cannot be cultured on artificial medium. Using *Galerina marginata*, an unrelated saprotrophic mushroom that grows and produces  $\alpha$ -amanitin in culture, we describe a method for producing  $^{15}\text{N}$ -labeled  $\alpha$ -amanitin using growth media containing  $^{15}\text{N}$  as sole nitrogen source. A key to success was preparing  $^{15}\text{N}$ -enriched yeast extract via a novel method designated “glass bead-assisted maturation.” In the presence of the labeled yeast extract and  $^{15}\text{N}$ - $\text{NH}_4\text{Cl}$ ,  $\alpha$ -amanitin was produced with >97% isotope enrichment. The labeled product was confirmed by HPLC, high-resolution mass spectrometry, and NMR.

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## 1. Introduction

Amatoxins, including  $\alpha$ -amanitin and  $\beta$ -amanitin, are hepatotoxic bicyclic peptides found in certain mushrooms, especially the genus *Amanita* (Wieland, 1986). Fatal poisonings occur every year throughout the world. The oral  $\text{LD}_{50}$  of  $\alpha$ -amanitin is approximately 0.1 mg/kg, and a lethal dose can be obtained by eating a single large basidiocarp (fruiting body) (Wieland, 1986).  $\alpha$ -Amanitin is also made by mushrooms in other genera, such as *Galerina* and *Lepiota* (Luo et al., 2012; Sgambelluri et al., 2014). The amatoxins and related phallotoxins are biosynthesized on ribosomes, unlike most other fungal cyclic peptides (Hallen et al., 2007; Luo et al., 2009, 2014).

Most cases of amanitin poisoning are accidental due to misidentification of wild mushrooms. In these cases, a history of mushroom consumption and clinical evidence of liver failure are

sufficiently strong indicators of amanitin poisoning to guide medical treatment. On the other hand, in some accidental or intentional poisonings the patient is not aware of having eaten mushrooms. Dogs and cats are also often victims of unobserved  $\alpha$ -amanitin-related mushroom poisonings (Puschner et al., 2007). There is also some concern that fungal toxins such as  $\alpha$ -amanitin have the potential to be used as bioweapons (Madsen, 2001; Russell and Paterson, 2006).

Stable-isotope labeled compounds have many applications in biochemical research, including metabolism and pharmacokinetics of drugs and other foreign compounds in animals and humans (Baillie, 1981; Haskins, 1982; Mutlib, 2008). One major use of stable-isotope labeled compounds is as internal standards to quantify metabolites in complex matrices such as blood, stomach contents, and urine. Stable-isotope labeled mycotoxins, such as fumonisin FB1 and deoxynivalenol, are used for the analysis of food products and animal feed (Hartl et al., 1999; Häubl et al., 2006; Spanjer et al., 2008; Malachová et al., 2014).

There are currently no available internal standards for any of the amatoxins (Johnson et al., 2010). To the best of our knowledge,  $\alpha$ -amanitin has never been chemically synthesized, which thwarts this route to producing  $\alpha$ -amanitin labeled with either radioactive

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or stable isotopes. Chemical radiolabeling with  $^{125}\text{I}$  led to a chemically distinct compound with reduced activity (Morris et al., 1978; Faulstich et al., 1981). In vivo labeling is hindered by the difficulty of culturing most species of *Amanita*. Even culturable species of *Amanita* grow very slowly (Zhang et al., 2005). *Galerina marginata*, on the other hand, is a saprotrophic mushroom that can be grown in culture (Enjalbert et al., 2004; Gulden et al., 2005). *G. marginata* contains  $\alpha$ -amanitin in its fruiting bodies and the mycelium produces it in liquid culture (Benedict et al., 1966; Benedict and Brady, 1967; Muraoka and Shinozawa, 2000). We studied this fungus as a possible route for the production of heavy-isotope labeled  $\alpha$ -amanitin that could be used as an internal standard for mass spectrometric analyses in forensic and clinical situations.

## 2. Materials and methods

### 2.1. $^{15}\text{N}$ -labeled yeast extract

Dried baker's yeast (*Saccharomyces cerevisiae*) was purchased at a local grocery store and grown on YPAD agar (1% yeast extract, 2% Bacto peptone, 2% dextrose, 2% Bacto agar, and 0.0075% L-adenine hemisulfate). For seed culture, a fresh colony of *S. cerevisiae* was transferred into 4 ml  $^{15}\text{N}$ -YNB medium, composed of (per liter) 30 g glucose, 1.2 g  $^{15}\text{NH}_4\text{Cl}$  (Sigma–Aldrich #299251, 98%  $^{15}\text{N}$ ), 1 g  $\cdot\text{KH}_2\text{PO}_4$ , 0.5 g  $\cdot\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 0.1 g  $\cdot\text{NaCl}$ , 0.1 g  $\cdot\text{CaCl}_2$ , 0.002 mg biotin, 0.4 mg calcium pantothenate, 0.002 mg folic acid, 2 mg inositol, 0.4 mg niacin, 0.2 mg p-aminobenzoic acid, 0.4 mg pyridoxine  $\cdot\text{HCl}$ , 0.2 mg riboflavin, 0.4 mg thiamine  $\cdot\text{HCl}$ , 0.5 mg boric acid, 0.04 mg copper sulfate, 0.1 mg sodium iodide, 0.2 mg ferric chloride, 0.4 mg magnesium sulfate, 0.2 mg sodium molybdate, and 0.4 mg zinc sulfate. The cultures were grown overnight with rotary shaking at 250 rpm and 30 °C. The seed culture was then transferred to 50 ml  $^{15}\text{N}$ -YNB medium in 250-ml baffled flasks and incubated under the same shaking conditions overnight. The next day, the culture was transferred to 500 ml  $^{15}\text{N}$ -YNB medium in 2-L baffled flasks and incubated under the same shaking conditions for 24 h. The culture was centrifuged at 10,000 g for 10 min, the pellet re-suspended in a minimum amount of double de-ionized water (starting with 1 ml) with constant stirring until the entire pellet was re-suspended, and then transferred into a 50-ml conical screw cap tube (Sarstedt). Resuspension typically required 3–5 ml water per liter culture.

The suspension was then subjected to a procedure designated “glass-beads-assisted maturation”. This method is based on a commercial process used for making yeast extract. On the first of three days, the suspension was vortexed without glass beads for 2 min once per hour for 6 h. On the second day, 1 ml of glass beads was added to every 3 ml of suspension followed by vortexing twice for 2 h each time, once in the morning and once 5 h later. The glass beads (Sigma–Aldrich #G8772) were 425–600  $\mu\text{m}$  in diameter and washed for 1 h with 1 N HCl, rinsed thoroughly with deionized water, and autoclaved. The same vortexing treatment continued on the third day without adding additional glass beads. The suspension was centrifuged at 10,000 g for 5 min and the supernatant transferred to a clean tube. The pellet was then washed twice with an equal amount of water by vortexing for 30 min. The three pooled supernatants were lyophilized, resulting in  $^{15}\text{N}$ -labeled yeast extract in powder form. The yield was typically 0.8–1.4 g  $^{15}\text{N}$ -yeast extract per liter culture.

### 2.2. Culture of *G. marginata* in $^{15}\text{N}$ -medium

Strain CBS 339.88 (obtained from the Centraalbureau voor Schimmelcultures, Utrecht, Netherlands) was maintained on standard potato dextrose agar (PDA) by transferring every two months.

This particular strain is monokaryotic and makes  $\alpha$ -amanitin on artificial media (Luo et al., 2012). The medium, designated  $^{15}\text{N}$ -HSV-5C (Muraoka and Shinozawa, 2000), contains, in 1 L, 1 g  $^{15}\text{N}$  yeast extract, 5 g glucose, 0.2 g  $^{15}\text{NH}_4\text{Cl}$ , 0.1 g KCl, 0.1 g  $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$ , 1 mg thiamine  $\cdot\text{HCl}$ , and 0.1 mg biotin, pH 5.2.  $^{15}\text{N}$ -HSV-5C (70 ml) in 250 ml flasks were used for liquid culture and inoculation was done by transferring 10 thin slices (8 mm  $\times$  8 mm  $\times$  1 mm) of the *G. marginata* PDA agar culture to each flask. Only the freshest edges of the mycelium were used. The slices were separated from each other with gentle swirling by hand. The flasks were then incubated at 110 rpm at room temperature for up to 40 d. Typically, *G. marginata* forms spherical mycelial balls up to 1.5 cm in diameter under these conditions and the color gradually changes from white to slightly brown (Fig. 1). A time course was done by collecting two mycelial balls from each flask every 3 days starting at day 20 with two biological replicates. In total, nine time points were obtained. The harvested mycelium was lyophilized to dryness and stored at  $-20$  °C.

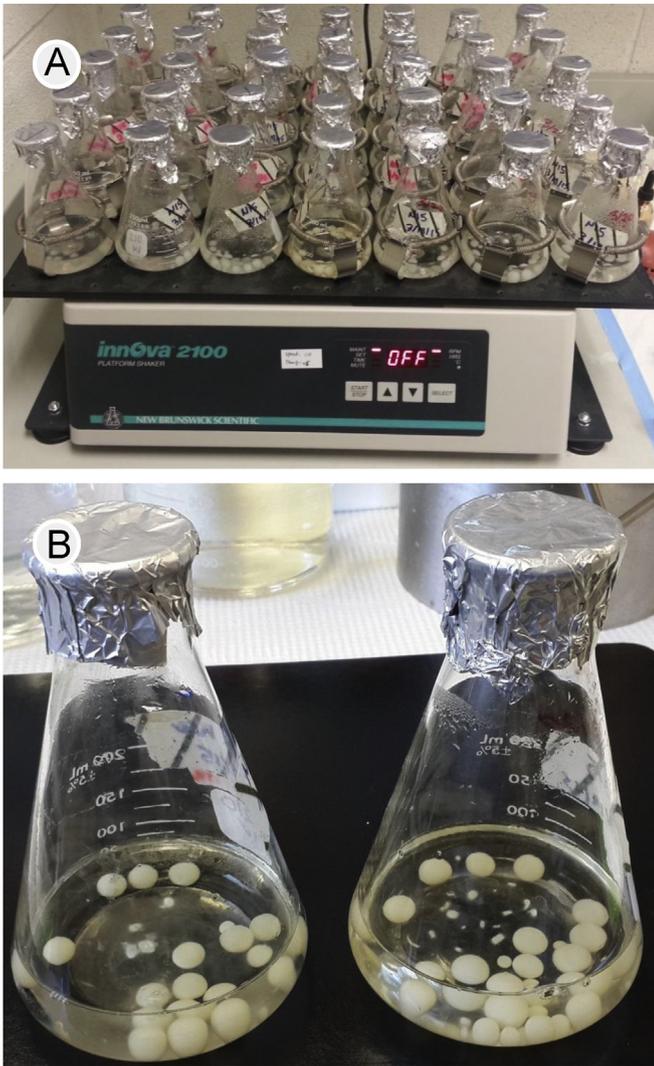
### 2.3. Toxin extraction

Lyophilized fungal mycelium (0.02 g for the time course experiments, or 0.5 g for large scale extractions) was ground in liquid nitrogen in a ceramic mortar and pestle. Extraction solution, composed of methanol:water:0.01 M HCl (5:4:1) (Enjalbert et al., 1992; Hallen et al., 2003), was added at 5 ml/0.1 g biomass. The suspensions were ground for another 5 min and then transferred to 1.5-ml microcentrifuge tubes (for the time course) or 50-ml conical screw-cap tubes (Sarstedt) (for the large scale extractions) and incubated at room temperature overnight. The next day, the suspensions were centrifuged at 10,200  $\times$  g for 3 min and the supernatants were collected, lyophilized, and re-suspended in 120  $\mu\text{l}$  water (for the time course) or 3 ml water (for the large scale extraction). The extracts were then boiled for 5 min in 1.5-ml microcentrifuge tubes sealed with clamps and centrifuged at 10,200  $\times$  g for 5 min. The supernatants were filtered through a 0.22  $\mu\text{m}$  filter (Millipore Millex-GV for the time course experiments or Millex-GS for large scale extractions) and stored at  $-20$  °C.

### 2.4. Toxin analyses and purification

The HPLC system was an Agilent Model 1200 equipped with a multiwavelength UV detector set to 280, 295, and 305 nm and an Agilent quadrupole 6120 mass spectrometer. Elution solution A was 0.02 M aqueous ammonium acetate:acetonitrile (90:10, v/v), adjusted to pH 5 with glacial acetic acid, and solution B was 0.02 M aqueous ammonium acetate:acetonitrile (76:24, v/v), also adjusted to pH 5. The flow rate and gradient profile were as described by Enjalbert et al. (1992).

For the time course experiments, separations were performed on a Higgins Proto 300 analytical C18 column (150  $\times$  4.6 mm, 5  $\mu\text{m}$  particle size, Higgins Analytical, Mountain View, CA). For large-scale toxin purification, a two-step chromatography procedure was used. The first separation was performed on a semi-preparative C18 column (25 cm  $\times$  10 mm, 5  $\mu\text{m}$ , Supelcosil LC-18 Semi-prep, Supelco, Bellefonte, PA). The flow rate was 2 ml/min with a step-wise gradient profile of 100% A for 3 min, 43% A for 7 min, and 0% A for 9 min. Loading was 1 ml of the toxin extract. Fractions were collected and 10  $\mu\text{l}$  of each sample submitted to high-resolution MS analysis on a Waters Xevo G2-S QToF system with all  $^{14}\text{N}$  replaced by  $^{15}\text{N}$  in the parameters setting. Fractions containing  $^{15}\text{N}$ -labeled  $\alpha$ -amanitin were pooled, lyophilized, and then re-dissolved in 100  $\mu\text{l}$   $\text{H}_2\text{O}$ . A second separation was performed on the Higgins C18 column. The flow rate was 1 ml/min with a gradient of 100% solution A to 100% solution B in 15 min. The fractions containing

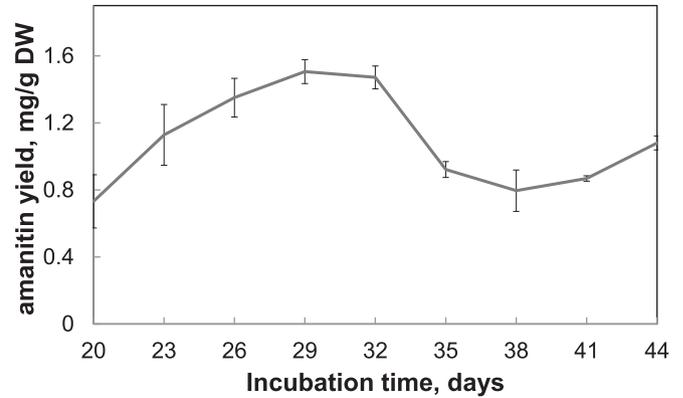


**Fig. 1.** Liquid culture of *G. marginata*. A. Shake culture. B. Close-up of flasks showing mycelial balls.

amanitin were collected, dried under vacuum (SpeedVac, Thermo Scientific), and re-dissolved in 100  $\mu$ l H<sub>2</sub>O. A third HPLC analysis was used to measure the purity of the resulting toxin. Purified toxin (100  $\mu$ l) was run under the same conditions as described in the second separation. Purity of the toxin was calculated by integration of the corresponding peak area at wavelength 280 nm. The  $\alpha$ -amanitin was quantitated by comparison to standard  $\alpha$ -amanitin (Sigma–Aldrich #A2263).

### 2.5. NMR of <sup>15</sup>N-labeled $\alpha$ -amanitin

<sup>15</sup>N-labeled  $\alpha$ -amanitin was dissolved in 90% H<sub>2</sub>O/10% D<sub>2</sub>O to a final concentration of 1.8 mM. 4,4-Dimethyl-4-silapentane-1-sulfonic acid (DSS) (10  $\mu$ M) (Sigma–Aldrich) was included as the chemical shift reference. A 2-D <sup>1</sup>H–<sup>15</sup>N-HSQC spectrum was recorded at 25 °C with 2048 data points in F2, 256 increments in F1, 16 scans per increment, on a Bruker Avance 900 MHz spectrometer using a TCI cryoprobe. Spectral windows were 16 ppm in F2 and 32 ppm in F1. <sup>15</sup>N shifts were referenced indirectly (Wishart et al., 1995).

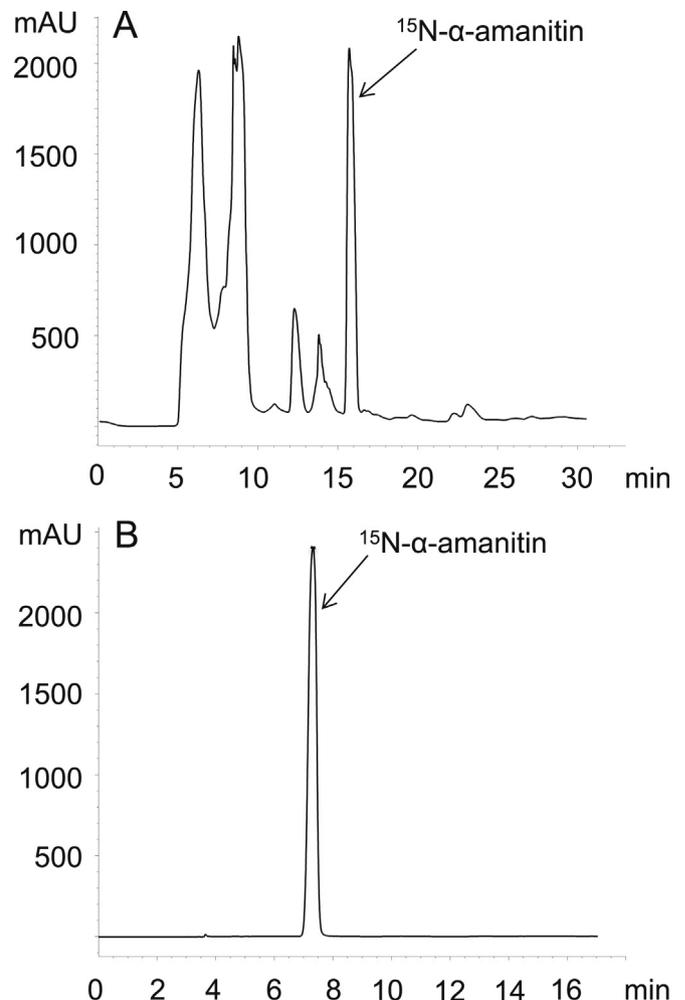


**Fig. 2.** Time course of <sup>15</sup>N-labeled  $\alpha$ -amanitin accumulation in *G. marginata* grown in shake culture. Each data point is the average of two biological replicates. Error bars indicate the range.

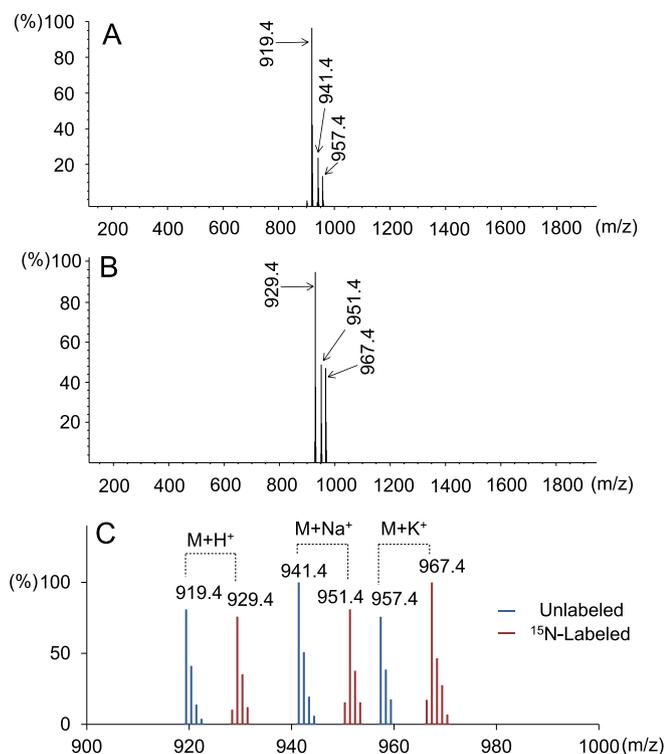
## 3. Results

### 3.1. Fungal culture and toxin production

In preliminary studies, *G. marginata* was found to grow poorly in the absence of yeast extract. We tested a variety of alternate



**Fig. 3.** HPLC chromatograms of the (A) first and (B) third separation of <sup>15</sup>N-labeled  $\alpha$ -amanitin. The y-axis is OD<sub>280</sub> (mAU).



**Fig. 4.** Mass spectrometric analysis of native and  $^{15}\text{N}$ -labeled  $\alpha$ -amanitin. A. Native amanitin.  $[\text{M} + \text{H}^+] = 919.4$ ,  $[\text{M} + \text{Na}^+] = 941.4$ , and  $[\text{M} + \text{K}^+] = 957.4$ ; B.  $^{15}\text{N}$ -amanitin.  $[\text{M} + \text{H}^+] = 929.4$ ,  $[\text{M} + \text{Na}^+] = 951.4$ , and  $[\text{M} + \text{K}^+] = 967.4$ . (C) Overlaid spectra of  $^{14}\text{N}$ -amanitin and  $^{15}\text{N}$ -amanitin, shown in expanded scale.

nitrogen sources (yeast nitrogen base,  $\text{NH}_4\text{Cl}$ , amino acids, and  $(\text{NH}_4)_2\text{SO}_4$ ), but none of them were satisfactory. Commercial  $^{14}\text{N}$  yeast extract was found to result in poorly labeled amanitin, i.e., the enrichment with  $^{15}\text{N}$  was only ~50%. A commercial source being unavailable, we prepared  $^{15}\text{N}$ -yeast extract in-house.

Using  $^{15}\text{N}$  yeast extract and  $^{15}\text{NH}_4\text{Cl}$  as the sole sources of N, toxin production by *G. marginata* grown in shake culture peaked at

about day 29 at 1.6 mg/g dry weight (Fig. 2). For large scale purification, the mycelial mass was harvested at day 29 or 30.  $\alpha$ -Amanitin extracted from lyophilized mycelium stored for 6 months at  $-20^\circ\text{C}$  showed no signs of degradation.

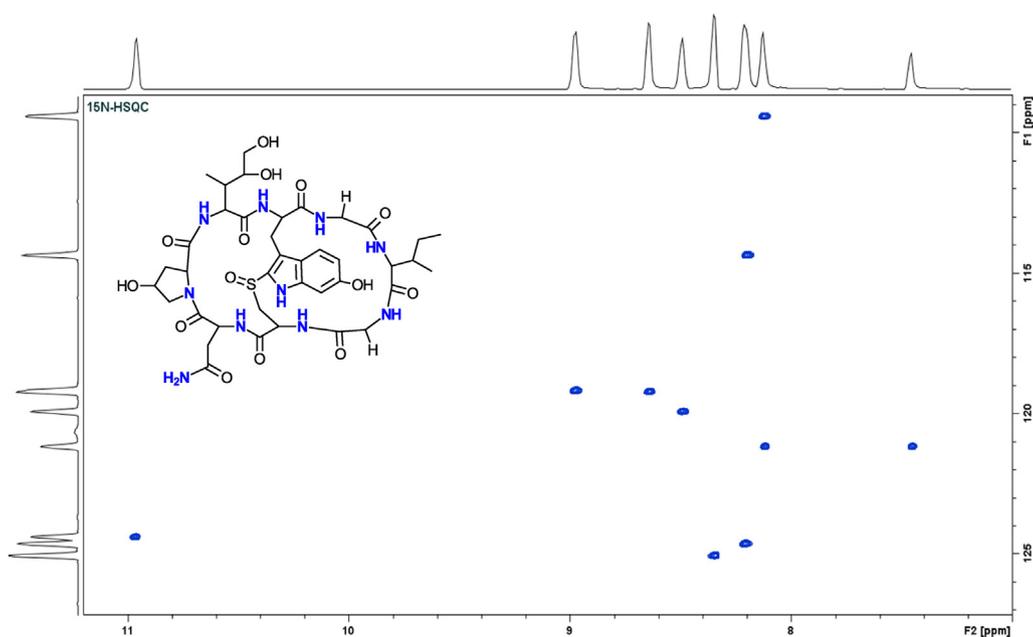
Toxin production was monitored by HPLC reverse-phase separation and UV detection. A fully  $^{15}\text{N}$ -labeled form of  $\alpha$ -amanitin (native mass 918.4) would have a mass of 928.4 due to the ten N atoms. Growing in liquid  $^{15}\text{N}$ -HSV-5C, *G. marginata* still produced a major peak of absorbance at 280 nm at the same retention time as native  $\alpha$ -amanitin, but the signal at  $m/z$  919.4  $[\text{M} + \text{H}^+]$  was no longer present. Instead, a new mass of 929.4  $[\text{M} + \text{H}^+]$  became the most significant, indicative of a compound labeled at all 10 N atoms with  $^{15}\text{N}$ .

### 3.2. Characterization of $^{15}\text{N}$ -labeled $\alpha$ -amanitin

Based on integration of the  $\alpha$ -amanitin  $\text{OD}_{280}$  peak at the last stage of purification (Fig. 3), the total amount of  $^{15}\text{N}$ -labeled  $\alpha$ -amanitin obtained was 8 mg from *G. marginata* harvested from approximately 6 l of culture. The chemical purity judged by integration at  $\text{OD}_{280}$  was estimated to be 99.8%. Based on the relative peak height of the  $m/z$  signal one mass unit smaller than the fully labeled compound (i.e., a  $\text{M} + \text{H}^+$  signal of 928.4), the isotope enrichment was calculated to be >97%. The MS spectra of native and  $^{15}\text{N}$ -labeled  $\alpha$ -amanitins are shown in Fig. 4.

$\alpha$ -Amanitin has a formula of  $\text{C}_{39}\text{H}_{54}\text{N}_{10}\text{O}_{14}\text{S}$  with a monoisotopic mass of 918.3541. The calculated mass of the  $[\text{M} + \text{H}^+]$  ion was 919.3620. The high resolution mass spectrum of purified  $^{15}\text{N}$ -labeled  $\alpha$ -amanitin showed a mass of 929.3330, matching the formula  $\text{C}_{39}\text{H}_{55}^{15}\text{N}_{10}\text{O}_{14}\text{S}$  (calculated mass 929.3323) with an error of 0.8 ppm.

The two-dimensional  $^1\text{H}$ - $^{15}\text{N}$  HSQC spectrum of the labeled product contained ten cross-peaks corresponding to the ten  $^{15}\text{N}$  nuclei in  $\alpha$ -amanitin (Fig. 5). Three resonances are readily assignable: A cross-peak with a  $^1\text{H}$  shift of 10.96 ppm can be assigned to the tryptophan indole NH and two other resonances at 7.45 and 8.12 ppm to the amide  $\text{NH}_2$  of the side-chain of asparagine. All ten signals were present at approximately the same intensity, suggesting equal incorporation of the  $^{15}\text{N}$  label at each position.



**Fig. 5.**  $^1\text{H}$ - $^{15}\text{N}$  HSQC spectrum of  $^{15}\text{N}$ -labeled  $\alpha$ -amanitin. The spectrum shows ten signals corresponding to the ten N atoms in  $\alpha$ -amanitin.

#### 4. Discussion

To the best of our knowledge, this is the first production of underivatized labeled  $\alpha$ -amanitin. A major impediment to development of a successful in vivo protocol using *G. marginata* was the need of this fungus for yeast extract to grow and produce amanitin. Commercial ( $^{14}\text{N}$ ) yeast extract caused excessive isotope dilution, and therefore we developed a novel method to make  $^{15}\text{N}$ -labeled yeast extract called “glass bead-assisted maturation”. Using this, we were able to produce  $^{15}\text{N}$ -amanitin with better than 98% labeling of its ten N atoms. The product is sufficient for use as an internal standard in mass spectrometry-based applications.

Unlike most *Amanita* species, *G. marginata* is a saprotrophic fungus and therefore can be more readily cultured than the obligately ectomycorrhizal amanitin-producing species of *Amanita* (Benedict et al., 1966, 1967; Zhang et al., 2005; Luo et al., 2012, 2014). In comparison, the other reported case of amanitin production in culture used the fungus *Amanita exitialis*, which produces  $\alpha$ - and  $\beta$ -amanitin on agar medium. The process takes up to 50 days and the amanitin levels reach ~10% of the concentration found in basidiocarps (Zhang et al., 2005). To the best of our knowledge, *A. exitialis* has not been used to produce labeled amanitins. *G. marginata* grows faster and reaches peak production of  $\alpha$ -amanitin around 30 d. The toxin concentration found in *G. marginata* culture was calculated to be two times that found in the culture of *A. exitialis*. In our lab, consistent toxin production has been observed using the same strain of *G. marginata* for over eight years, suggesting that its biosynthetic pathway is stable.

#### Acknowledgments

This research was supported by contract 200-2014-M-58133 from the Centers for Disease Control, Atlanta, GA. Additional support came from grant 1R01-GM088274 from the U.S. National Institutes of Health General Medical Sciences. We thank the Michigan State University (MSU) Mass Spectrometry Core Facility for high-resolution MS, and the MSU Max T. Rogers NMR facility for NMR analysis. We thank Yair Shachar-Hill, Department of Plant Biology, MSU, for advice on in vivo isotope labeling, and Melissa Carter, CDC, for valuable discussions about the project.

#### Transparency document

Transparency document related to this article can be found online at <http://dx.doi.org/10.1016/j.toxicon.2015.06.018>.

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